



FormaPure RNA:

Extended Protocol for RNA Isolation from FFPE Sample

Refer to www.beckmancoulter.com/ifu for updated protocols. For questions regarding this protocol, call Technical Support at Beckman Coulter at 1-800-369-0333.

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Introduction

The FormaPure RNA extraction and purification kit uses the patented Beckman Coulter SPRI paramagnetic bead-based technology to isolate RNA from formalin-fixed, paraffin-embedded (FFPE) tissue without the use of xylene. This kit has been optimized for use with downstream sequencing and PCR-based assays. Specifically, RNA isolated using the FormaPure RNA kit is compatible with the following downstream applications:

- RNA-seq
- Endpoint or qRT-PCR

FormaPure RNA isolates RNA from tissue sections totaling a thickness of up to $3 \times 10 \mu\text{m}$. The protocol can be performed in both 96-well plates (manually and automated) and in 1.5 mL tubes (manually only). Nucleic acid extraction begins with the solubilization of the paraffin from the tissue slices in tubes. An enzymatic lysis step digests the tissue and releases the nucleic acids, as well as gently decrosslinks RNA. The remaining protocol can be carried out in plates or tubes. A binding solution is added to immobilize the nucleic acids to the surface of the SPRI beads. Contaminants are rinsed away using a simple washing procedure, DNA is removed from the sample and RNA is again immobilized on the surface of the SPRIbeads before elution with water.

Kit Specifications

Kit Type	Part Number	Number of Preps
Medium	C19158	96
Small	C19157	50

Warnings and Precautions

Read and observe the following safety information.

IMPORTANT The  symbol indicates a potential safety risk involving the material, action, or equipment required for executing a procedural action; when you see the  symbol, return to this section to review relevant safety information.

	DANGER
Proteinase K	
H315	Causes skin irritation.
H319	Causes serious eye irritation.
H334	May cause allergy or asthma symptoms or breathing difficulties if inhaled.
H335	May cause respiratory irritation.
P261	Avoid breathing vapors.
P280	Wear protective gloves, protective clothing and eye/face protection.
P284	In case of inadequate ventilation, wear respiratory protection.
P304+P340	IF INHALED: Remove person to fresh air and keep at rest in a position comfortable for breathing.
P342+P311	If experiencing respiratory symptoms: Call a POISON CENTER or doctor/physician.
	Safety Data Sheet is available at techdocs.beckmancoulter.com .

CAUTION

Risk of chemical injury from Proteinase K. To avoid contact with Proteinase K, wear appropriate personal protective equipment, including protective eyewear, gloves, and suitable laboratory attire. Refer to the Safety Data Sheet for details about chemical exposure before using the chemical.

 **CAUTION**

Risk of burning from hot liquid splattering into your eyes or onto your skin. Wear appropriate personal protective equipment while incubating the samples. Place tube cap locks on the tubes to prevent the tops of the tubes from opening during incubation.

Materials Supplied

The following reagents are supplied in the FormaPure RNA kit. The reagent icon, which is located on the top of the corresponding bottle, is included in the instructions as a visual aid to ensure the correct reagent is used.

NOTE Refer to the product labels for expiration dates.

Reagent	Icon	Storage Conditions
Mineral Oil		15 to 30 °C
Lysis		15 to 30 °C
Bind		15 to 30 °C
Re-Bind		15 to 30 °C
Proteinase K	-	15 to 30 °C

Materials Required but not Supplied

FormaPure samples can be processed in a 96-well plate or tube format. Refer to the tables below for the items required for this procedure:

- *Hardware & Accessories*
- *Consumables*
- *Reagents*

Hardware & Accessories

Item	Type
Adjustable Heat Source	<ul style="list-style-type: none"> Thermomixer with 1.5 mL tubes and plate adaptor and heated lid <p>Or</p> <ul style="list-style-type: none"> Hybex with 1.5 mL tubes and plate adaptor <p>Two heat sources of any type are recommended for the protocol.</p>
Vortexer	Not specified.
Microcentrifuge	Beckman Coulter Microcentrifuge 16 Or equivalent.
Bead Separation Magnet	<ul style="list-style-type: none"> Agencourt SPRIStand Magnetic 6-Tube Stand (for 1.5, 1.7, or 2.0 mL tubes) (Beckman Coulter), PN A29182 <p>Or</p> <ul style="list-style-type: none"> V&P Scientific 7 Bar Magnet, PN VP 771MWZM-1ALT (for 96-Well Plate)

Consumables

Item	Type
Microcentrifuge Tubes	1.5 mL
Cap Locks	for Microcentrifuge Tubes
96-Well Plate	1.2 mL, ThermoFisher Scientific, PN AB1127 Or equivalent.
96-Well Storage Plate	200 µL
PCR Adhesive Seals	for 96-Well Plate

Reagents

Item	Supplier	Catalog Name	Catalog Number
100% Ethanol (Molecular Grade) ^a	AmericanBio	Ethanol, Absolute Alcohol, 200 Proof, Anhydrous	AB00138
DNase I	ThermoFisher Scientific	Ambion DNase I (RNase-free)	AM2222 or AM2224
Nuclease-Free Water ^a	ThermoFisher Scientific	Nuclease-Free Water (not DEPC-Treated)	AM9932

a. The recommended **Supplier**, **Catalog Name**, and **Catalog Number** for this item is provided; if necessary, an equivalent product may be substituted for the listed product.

Process Overview



1. Deparaffinization at 80°C.
2. Lysis/decrosslink at 60°C.
3. Bind.
4. 80% Ethanol wash.
5. DNase I treatment.
6. Re-Bind.
7. 80% Ethanol wash.
8. Elution Buffer.
9. Transfer.

Protocol for RNA Isolation

Before You Begin

- Preheat adjustable heat sources to 80°C and 60°C.
- Prepare 80% **Ethanol** from 100% stock using **Nuclease-Free Water**.
IMPORTANT Do not use a previously-prepared solution, as it may have a lower ethanol percentage, causing yield loss.
IMPORTANT This protocol uses ethanol in multiple steps. Dispose of supernatant containing ethanol waste in accordance with local regulations and acceptable laboratory practices.
- Wear appropriate personal protective equipment (PPE) when handling samples and reagents.

Procedure

1 Sample Preparation:

For each sample, transfer one to three **10 µm** FFPE tissue sections into a 1.5 mL tube.

2 Deparaffinization:

- a. Add **450 µL** of **Mineral Oil** (MO) to each sample and immerse the sections completely with a pipette tip.

NOTE Make sure that the sample is completely immersed and does not float due to attached bubbles.

- b.  Incubate at 80°C for 5 minutes.
- c. After incubation, vortex the tubes two times, for five seconds each time, to solubilize the paraffin and disperse the tissue.

3 Tissue Digestion:

- a. Add 200 µL of Lysis  to each sample.
- b. Centrifuge the tubes at 10,000 × g for 15 seconds. The mineral oil forms a separate upper phase.
NOTE  Incubate the tubes for 3 more minutes at 80°C if the mineral oil layer appears cloudy and the tissue is stuck at the interface of mineral oil and lysis buffer. After the incubation, make sure to cool the tubes for 2 minutes before adding Proteinase K.
- c.  Add 20 µL of Proteinase K to the aqueous, lower phase and mix by pipetting up and down 10 times without disrupting the upper phase.
- d. Incubate the tubes at 60°C for 120 minutes.

4 Lysate Transfer:

- a. Take the tubes out of the heat source and centrifuge the tubes at 10,000 × g for 5 minutes.
- b. Transfer all of the clear lysate (lower phase) to a 96-well plate, or to 1.5 mL tubes, without disrupting the upper phase (mineral oil) or the pellet.
NOTE If the tissue is clogging the pipette tip, you may centrifuge the tubes for additional time.
NOTE Minimize the amount of Mineral Oil that is transferred along with the lysate. However, a small amount of Mineral Oil carryover does not affect downstream applications.

5 First Bind:

- a. Fully re-suspend the Bind  by shaking or vortexing.
- b. Add 300 µL of Bind  to each sample and mix by pipetting up and down 10 times with a P1000 pipette set at 350 µL. Mix gently to minimize the generation of bubbles.
- c. Incubate at room temperature for 5 minutes.

- d. Place the samples on the magnet for 10 minutes; if the solution is not fully clear after 10 minutes, incubate until clear. If using:
 - **A 96-well plate**, place the samples on the bar magnet plate.
 - **Tubes**, place the tubes in an Agencourt SPRIStand Magnetic 6-Tube Stand.
- e. With the samples on the magnet, aspirate the supernatant without disrupting the beads. Discard the supernatant.

6 Ethanol Wash:

- a. Remove the samples from the magnet.
- b. Add **750 μ L** of freshly prepared **80% Ethanol** to each sample.
- c. Using a P1000 pipette set at 600 μ L, mix by pipetting up and down 20 times, or until the beads are fully re-suspended.
- d. Place the samples on the magnet for 3 minutes; if the solution is not fully clear after 3 minutes, incubate until clear.
- e. With the samples on the magnet, aspirate the supernatant without disrupting the beads. Discard the supernatant.
- f. Air dry the samples on the magnet for 10 minutes.

7 DNase I Treatment:

- a. Remove the samples from the magnet.
- b. Add **80 μ L** of **Nuclease-Free Water** to each sample.
- c. Add **10 μ L** of **10 \times DNase I buffer** and **10 μ L** of **DNase I** to each sample.
- d. Mix by pipetting up and down 5 times with a P200 pipette set at 80 μ L to thoroughly distribute the buffer and enzyme. Mix gently to minimize the generation of bubbles.
- e. Cover the plate with an adhesive seal, or close the tubes, and incubate at **37°C** for **20 minutes**.

8 Re-Bind:

- a. Add **150 μ L** of **Re-Bind** **RBA** to each sample and mix by pipetting up and down 10 times with a P200 pipette set at 150 μ L. Mix gently to minimize the generation of bubbles.
- b. Incubate at room temperature for **5 minutes**.
- c. Place the samples on the magnet for 10 minutes; if the solution is not fully clear after 10 minutes, incubate until clear.
- d. With the samples on the magnet, aspirate the supernatant without disrupting the beads. Discard the supernatant.

9 Ethanol Wash:

- a. Remove the samples from the magnet.
 - b. Add **750 μL** of freshly prepared **80% Ethanol** to each sample.
 - c. Using a P1000 pipette set at 600 μL , mix by pipetting up and down 20 times, or until the beads are fully re-suspended.
 - d. Place the samples on the magnet for 3 minutes; if the solution is not fully clear after 3 minutes, incubate until clear.
 - e. With the samples on the magnet, aspirate the supernatant without disrupting the beads. Discard the supernatant.
 - f. Air dry the samples on the magnet for 10 minutes.
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10 Elution:

- a. Remove the samples from the magnet.
 - b. Add a minimum of **40 μL** of **Nuclease-Free Water** to each sample and mix by pipetting up and down 10 times, or until beads are fully re-suspended, with a P200 pipette set at 30 μL .
 - c. Cap tubes or cover the plate with a PCR adhesive seal and incubate at **60°C** for one minute.
 - d. Place the samples on the magnet for 1 minute; if the solution is not fully clear after 1 minute, incubate until clear.
 - e. With the samples on the magnet, transfer as much of the supernatant as possible to a 96-well storage plate, or to a new tube, without disturbing the magnetic beads.
 - f. Store at -20°C, or -80°C for long-term storage.
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Product Availability

FormaPure RNA

REF C19157 — FormaPure RNA, 50 Prep Kit

REF C19158 — FormaPure RNA, 96 Prep Kit

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For additional information, or if damaged product is received, call Beckman Coulter Customer Service at 800-742-2345 (USA or Canada) or contact your local Beckman Coulter Representative.

Glossary of Symbols is available at techdocs.beckmancoulter.com (PN C05838).



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